Centers of Motion Associated with EF-Tu Binding to the Ribosome

by

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Abbreviations and acronyms:

ASF: A-site finger

GAC: GTPase Associated Center

LSU: large subunit

rRNA: ribosomal RNA

SRL: sarcin ricin loop

SSU- small subunit

Abstract

Structural centers of motion (pivot points) in the ribosome have recently been identified by measurement of conformational changes in rRNA resulting from EF-G GTP hydrolysis. This series of measurements is extended here to the ribosome's interactions with the cofactor EF-Tu. Four recent EF-Tu bound ribosome structures were compared to unbound structures. A total of sixteen pivots were identified, of which four are unique to the EF-Tu interaction. Pivots in the GTPase associated center and the sarcin-ricin loop omitted previously, are found to be mobile in response to both EF-Tu and EF-G binding. Pivots in the intersubunit bridge rRNAs are found to be cofactor specific. Head swiveling motions in the small subunit are observed in the EF-Tu bound structures that were trapped post GTP hydrolysis. As in the case of pivots associated with EF-G, the additional pivots described here are associated with weak points in the ribosomal RNA structures such as non-canonical pairs and bulge loops. The combined set of pivots should be regarded as a minimal set. Only several states available to the ribosome have been presented in this work. Future, precise crystal structures in conjunction with experimental data will likely show additional functional pivoting elements in the ribosomal RNA.

Introduction

The ribosome is responsible for the dynamic process of translation.[1] It is comprised of two subunits, each consisting of RNA and protein. In Bacteria, the major co-factors that facilitate this process are the elongation factors EF-Tu and EF-G, initiation factor IF-2, and the release factor RF-3.[2] EF-G is thought to coordinate and hasten accommodation and translation of the tRNA by cycles of conformational rigidity and relaxation before and after GTP hydrolysis.[3] EF-Tu is believed to control tRNA mobility by disallowing incorrect codon-anticodon interactions.[4]

With these cofactors and others, the process of translation exhibits multiple motions including tRNA translocation, intersubunit ratcheting, and small subunit head swivel.[3, 5-7]

The motions of tRNA during the various stages of translation, including accommodation, are largely associated with reorientations of a structurally weak pivoting element.[5,8-10] Motion also exists in the mechanisms of intersubunit ratcheting and 30S head swiveling, which have previously been analyzed using high-resolution crystal structures,[6,11] cryo-EM structures,[12] and computational studies.[7,13,15] Recent high-resolution crystal structures of EF-G[16,17] and EF-Tu[18-21] associated ribosomes now allow further characterization of the cofactor dependent elements in the ribosomal RNA core.

Major pivoting elements associated with EF-G functionality were reported previously. [22] Herein, this effort is extended to identify pivoting positions associated with EF-Tu function in *Thermus thermophilus*. This is accomplished by comparison of four high resolution crystal structures of ribosome subunits bound and unbound to the cofactor EF-Tu.[18-21,23] In two bound structures the GTP is not hydrolyzed, while in the other two it is. The motions made obvious by alignment of the different structures are tabulated by the resulting greatest interhelix distance in Angstroms. When combined with the earlier EF-G results, a set of elements allowing large scale motion is identified in the rRNAs of *T. thermophilus.* The differences in the mobility of the described set of rRNAs hint at previously unreported functional differences between the two cofactors.

Results

Consistent with earlier studies,[22,24] partial overlap exists between pivoting elements associated with EF-Tu binding and those previously found to be associated with EF-G. Three

74 categories of pivots were recognized. This includes those that were active with both EF-Tu and
75 EF-G, those that are only mobile in the presence of EF-Tu, and those that are associated only
76 with EF-G. Table 1 and 2 summarize the average results for four individual comparisons.
77 Detailed results for the individual comparisons are provided in the Supplemental materials as
78 Tables S1-S8. Individual results are tabulated for pre-and post-GTP hydrolysis in Table S9 for
79 16S rRNA and Table S10 for 23S rRNA.

80 A total of 12 pivots are mobile in both sets of structural comparisons. In the SSU, these
81 are helices h6, h8, h33, h39, h40, and h44- all of which are associated with intersubunit bridges.
82 In the large subunit, these are the A- site finger H38, the b/L12 stalk H42, bridge b1a H69, the
83 uL1 stalk H76, as well as GTPase associated center (GAC), helix H89, and sarcin ricin loop
84 (SRL) helix H95. This set of elements is involved in cofactor binding and the tRNA translocation
85 process.

86 Four new pivoting elements specific to EF-Tu binding were found. These were helices
87 h14 and h17 in the small subunit and helices H10 and H59 in the large subunit A number of
88 pivots were active in the EF-G bound structures are inactive in the EF-Tu structures. In the
89 LSU, these are intersubunit bridge helices H34, H69, and H84. In the SSU, EF-G specific pivots
90 are in helices- h21, h26, h28, h31, h32, h36, h37, h41, h42, and h43. These include pivots
91 associated with the head swiveling motion initiated through EF-G-GTP hydrolysis and
92 intersubunit ratcheting.

93 Helices H69 and h28 both showed dependency on GTP hydrolysis by EF-Tu. Large scale
94 motion was observed in structures stalled by a GTP analogue in the pre-GTP hydrolysis state,[19]
95 while only smaller motions were observed in structures stalled by kirromycin.[18,20,21] Structures
96 2WRN, 2WRO (4V5G)[18] were trapped in the EF-Tu-GTP hydrolysis transition state with

kirromycin and paramomycin. Structures 2XQD, 2XQE and (4V5L)[19] were captured by GDPCP in the pre-hydrolysed state. Structures 2Y10, 2Y11 (4V5R, now superseded by 4V5S)[20] and 4ABR, 4ABS (4V8Q)[21] were captured in the post- GTP hydrolysis state with kirromycin. In the small subunit, h33 moves with respect to helix h28 alignment in pre-GTP hydrolysis structures 2WRN and 2XQD. The motion measured is 5.4 Angstroms and 5.8 Angstroms respectively. A ~4Angstrom full head swivel is seen as a result of the same alignment in the post-GTP-hydrolysis structures, 2Y10 and 4ABR. Helix H69 moved by 2.2 and 2.3 Angstroms in the post GTP hydrolyzed structures 2Y11, and 4ABS and more dramatically by 4.3, and 6 Angstroms before GTP hydrolysis in structures 2XQE and 2WRO.

Helices h10 and h42 both met the cutoff only once and with an overall average below 2.5 Angstroms were not considered to be mobile by the criteria used here. Helix h8 failed to meet the cut off in only one comparison and based on overall average is included as mobile. Finally, helices h43 and h17 in two cases exhibited modest mobility, while in the other two cases they showed essentially none. Figures 1 and 2 show the location of these three categories of pivots in the context of the bacterial rRNA secondary structure. The local context of the unique EF-Tu pivots in the 50S subunit is shown as an insert on Figure 1 and in higher resolution in Supplemental Figure S1.

Discussion

Large Subunit

Local motions resulting from cofactor association, intersubunit bridging, and the 30S head swivel play a large role in translation. [3,5-7] With respect to cofactor binding, both EF-G and EF-Tu contact the ribosome primarily at the GTP associated center (GAC), which includes

helices H43 and H44.[24] Though structurally similar, the two cofactors are thought to have a distinct binding mechanism.[24] An important feature related to cofactor selectivity in the GAC is the distance between the GAC and the sarcin ricin loop (SRL)[24] as well as the incoming cofactor P-loop.[25] The measurements obtained here allow up to 10 Angstroms of motion for the SRL which was previously described as immobile.[24] Further, the SRL fits the profile of a typical pivoting element, which includes a U-G wobble base pair that closes a three way junction. This structure likely allows the SRL the flexibility to accommodate the incoming cofactor.

In response to EF-Tu binding, motion is again seen in the tRNA and a series of pivoting elements around the tRNA extending from the A to the E site (Figures 1 and 3 and Figure S1). H76- the uL1 stalk, H38- the A-site finger (ASF) and H42- the bL12 stalk are found to be mobile. The ASF and the uL1 stalk contact the tRNA during translocation while H42 forms a series of functional contacts with the elements of the GAC.[25] The E-site- tRNA interaction lies directly upstream of the H82 stem as shown in supplemental Figure S2.

H82 is in direct proximity to H68, which has an internal bulge motif that suggests mobility and is known to contact the mobile tRNA in eukaryotes.[26] Helix H68 in turn, is in contact with H76- the uL1 stalk, predicted to move as it guides the tRNA towards the exit site. The uL1 stalk contacts the tRNA at residue G2112, but not G2116 as was predicted previously,[27] at least not in the 4V5L structure. It appears that the tRNA is accommodated throughout the PTC by a set of highly mobile elements- functionality known for the uL1 stalk but unreported in helices H68 and H82.

Helices H10 and H59 are uniquely mobile in the EF-Tu bound ribosome as shown in the supplemental materials in Figures S3 and S4. Helix H10 is another mobile element exclusive to the EF-Tu bound ribosome structure. Mobility in H10 can be explained, by its proximity i.e.

potential contact with proteins bL9 and bL28, which form a base for the highly mobile H76 that comprises the uL1 stalk bL28, is required for ribosome assembly.[28] Helix H59 is exclusively perturbed by EF-Tu. H59 is also a known contact site for the signal recognition particle (SRP).[29, 30] The Alu domain of the SRP mimics elongation factor structure in the PTC thereby arresting elongation by competition with elongation factor binding on the ribosome.[30] The 'minor-saddle motif' of the Alu domain is a flexible three way junction closed by a base pair mismatch, which-fits the general trend of structures that form pivoting elements.[30]

Intersubunit Bridging

In structures bound to EF-Tu, H69 is activated weakly, primarily in the post GTP-hydrolysis state. This mobility is related to the intersubunit rotation of the ribosome through bridge B2a and h44 of the SSU.[22] The combination of the small subunit h28 directed head swiveling motions and h69 mobility as a result of EF-Tu GTP hydrolysis are further evidence for a connected network of cofactor dependent pivoting rRNA elements.

H34 and H84 are also less mobile post EF-Tu binding. This may mean that these bridges are less utilized during the EF-Tu binding event and more heavily used in the intersubunit ratcheting process. Bridge B8 is also found to be highly mobile in the EF-Tu bound structure, while relatively inactive in the EF-G bound ribosome.

Small Subunit

Both helices h14 and h8, connected to bridge B8 in the small subunit are found to move ~3A in the current structure set (Figure 4). Although h14 is a relatively small helix, it exhibits

some conformational freedom in EF-Tu structures as a result. Proximal to this region (Figures S5 and S6) are mobile helices h6- the spur, h10, and h17.

Helix h44 moves by ~10 Angstroms in the EF-Tu bound ribosome vs. the unbound molecule. h44 contacts h8 and induces a 3.0 Angstrom change in the final residue of h8 which moves 2.1 Angstroms closer to h14 as shown in Figure 3. As a consequence h14 and h17 now show motion that was not seen in EF-G structures.

In addition to its contact with h8, h44 also contacts the base of h28, which is thought responsible for the majority of the head swivel motion.[15,22] Alignment of h28 and h32 in the small subunit results in motion of helix h33, which moves with respect to alignment at both helices in all four structures. This is expected as the result of mobility of the B1a bridge-(ASF, S13, S19), which forms contacts with helix 33.[31] Head swiveling of approximately 4 Angstroms is seen after alignment of pre GTP hydrolysis EF-Tu bound ribosomes. The swiveling motion is not as robust as that seen in the GTP hydrolyzed EF-G bound ribosome, but is still significant. This is perhaps surprising as only cofactor EF-G is heavily associated with the propagation of intersubunit ratcheting and consequently head swiveling motions.[3, 22]

Conclusions

A network of flexible rRNA interactions described earlier[22] proceeds through a series of motions when associated with elongation factors EF-Tu and EF-G. The two cofactors may be producing unique interaction sets. The EF-Tu set involves intersubunit bridge b8 in place of helix H69 of the LSU. Ratcheting and head swiveling motions are activated as a response to EF-G GTP hydrolysis but are weakly activated as a response to EF-Tu binding in agreement with earlier findings[22]. Another point of interest is EF-Tu's potential competition with the SRP protein

which binds helix H59 in the large subunit, an interaction missing from the EF-G protein. Overall, the results highlight the importance of weak sites in RNA structures in providing function and flexibility. This is likely to occur in other natural or synthetic RNAs. Indeed, the presence of a bulge or non-standard pair is very likely to be a site of significant flexibility in any RNA.

Materials and Methods

The PyMOL Molecular Graphics System, Version 1.7.6.2 Schrodinger, LLC (https://www.pymol.org/) was used to measure differences in crystal structures of ribosomal subunits, which are bound and unbound to EF-Tu. All structures were obtained from the PDB[32] (http://www.rcsb.org).The pivoting elements were identified by a global structural superposition followed by a local superposition as detailed previously[22]. Local sequences, which retained the greatest change after the global alignment, were then manually selected and aligned at the chosen stem sequences. In brief, the local superposition, the 'align' command in PyMOL was used on rigid "stem" sequences of mobile rRNA using available scripts.[33] Mobility here is defined as large scale motion (at least 2.5 Angstroms) post local alignment of cofactor bound ribosomes to cofactor unbound ribosomes. The cut off range for a pivot is thus 2.5 Angstroms, deemed reasonable against the highest resolution structures compared, All "major pivots" mentioned herein meet this requirement. This method forces a minimal average root mean square distance between all atoms of the stem sequence. Measured motion at the end of the helix is the result of the stem alignment and consequent change at the pivoting position. Single Watson–Crick matches were found suitable as alignments stems as they would yield the superposition of at least 30 atoms- enough to generate reproducible directionality. Though measurements made using this

method are relative (choice of alignment affects the magnitude of motion somewhat) the process consistently highlights elements shown to be mobile in experimental studies. Further, because only local stem alignments are used for measurement, observed changes are separated from global conformational changes of the ribosome. However, some flexible helices such as h34 and H68 are not readily detectable because no meaningful stem sequence is adjacent to the pivoting element.

A series of structures were compared in *T. thermophilus* using structures 2J01, 2J02 (4V51)[23] and 2WDI, 2WDG (4V5C)[34] as references. The global alignment of the standard structure sets showed an RMSD of 0.432 for the 16S rRNA and 0.345 for the 23S rRNA after removal of all non-rRNA structures. Standard structures 2J01 and 2J02 were first compared against EF-G bound structure pairs 4JUW, 4JUX (4V9H)[16]. In this case, the RMSD values were 1.951 for the 16S rRNA and 0.911 for the 23S rRNA far exceeding the background cutoff values as did all the other comparisons undertaken. PDB files 2WRI, 2WRJ (4V5F) were also compared[17]. Finally, standard structures 2J00, 2J01 were compared against 4 EF-Tu bound structures 2WRN, 2WRO (4V5G), 4ABR, 4ABS (4V8Q), 2XQD, 2XQE (4V5L), and 2Y10, 2Y11 (4V5R). [18-21] The structure from PDB set 4V5L is trapped in in a state prior to GTP hydrolysis, while structures 4V5R and 4V8Q are trapped in a post-GTP hydrolysis state. Although structure 4V5G is described as immediately after GTP hydrolysis, the EF-Tu domain conformation is thought to be similar to the pre-GTP hydrolysis conformation.[18] Results from this structure were thus averaged with those of structure 4V5L.

A potential problem with the method is areas of disorder in the compared crystal structures. Large B-factor regions may very well yield false positives in the identification of mobile rRNAs. However, high B value areas were not discounted as erroneous, because flexible

RNA is likely to yield crystals which are inherently disordered. To address this issue, published crystal structures, produced by various crystallization protocols were compared as described above. These consistently showed similar mobility as a result of cofactor binding, and in full agreement with published literature. Finally, it should be noted that the set of pivots is a minimal set representing the major points of flexibility. To this end, an average minimal motion of 2.5 Angstroms in the four comparisons considered was required for a pivot to be considered significant. Borderline cases likely exist such as h10 and h42 which were considered here and a change in the criterion would therefore reveal additional prospects.

Disclosure of Potential Conflicts of Interest

No potential conflicts of interest are disclosed

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Supplemental Material

Supplemental data and figures for this article can be accessed on the publisher's web site

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Figure Captions

Figure 1. Large subunit map with pivoting positions highlighted. Mobile pivots are highlighted: EF-Tu alone (blue), EF-G alone (green), both (red). The upper structural insert shows a selection of pivoting helices proximal to the tRNA as it moves towards the exit site. The lower insert shows H10 (orange) positioned to interact with proteins bL9 and bL28, which in turn contact the highly mobile H76/ uL1 stalk.

Figure 2. Small subunit secondary structure map with pivoting positions highlighted. Mobile pivots are highlighted: EF-Tu alone (blue), EF-G alone (green), both (red). Two additional pivots proposed previously[15] are labeled in purple. Helix h10, which is not considered to be a pivot as discussed in the text is labeled in black. The two EF-Tu specific pivots are in close proximity as shown in the insert. Helix h44 (black), activates bridge B8 helices h8 (blue) and h14 (red), in the small subunit. Mobile helices h6 (green)- the spur, h10 (pink), and h17(orange)-are also proximal to bridge B8.

Figure 3. Detailed examination of regions proximal to the tRNA as it moves towards the exit as shown in the upper insert of Figure 1. The helices may be accommodating the tRNA during translation. Helix 76 is known to guide the tRNA through continuous ionic interaction throughout the process. Helices H84 and H69 are found to be less mobile in EF-Tu bound ribosomes than EF-G bound ribosomes. The A, P, and E site tRNAs are shown in blue. EF-Tu is in green. Pivoting bases are shown as stick models. The structures compared (4V51 black and 4V8Q colored) are pre and post EF-Tu binding.

Figure 4. Bridge B8 in the SSU. Helix 44 moves by ~10 Angstroms in the EF-Tu bound ribosome vs. the unbound molecule. The unbound state is shown in black for each helix with the bound state in a unique color for each protein. Pivoting bases are shown as stick models. Helix h44 contacts h8 and induces a 3.0 Angstrom change in the final residue of h8 which moves 2.1 Angstroms closer to h14. Although h14 is a relatively small helix, it exhibits some conformational freedom, as a result. Helix 10, in proximity to the pivots, displays minor motion and connects h17 to bridge B8 as shown in supplemental Figures S5 and S6. Overall, the scheme shows a connected network of interactions, which explain partially, the structural consequence of EF-Tu ribosome association. Structure 2J00 is in black and blue. Structure 4ABR is in red and green)