

# Evolution of Resistant Mutants in Antibiotic Treated Bacterial Cultures

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## BACKGROUND

- Persistence is a rare non-genetic, non-heritable adaptive strategy that comes from phenotypic variants of bacteria, which enter a dormant-like state (2).
- Persister cells are an important health concern because they underlie the proclivity of recurrent infections to relapse, and they can serve as a reservoir from which drug resistance mutants can emerge (1).
- Despite reduced physiological activity, persister cells maintain their viability by sustaining a minimum adenylate energy charge, synthesizing spontaneously denatured or degraded proteins, and repairing antibiotic-induced DNA/protein/lipid damage.

## OBJECTIVES

- We approach the problem of enriching cultures with antibiotic resistant bacteria by treating the samples with antibiotics for 22 consecutive days.
- When Ofloxacin is introduced into the bacterial culture, the bacterial cells' DNA is damaged. Bacterial cells then have to respond to this damage by inducing SOS response genes (3).
- DNA repair causes random mutations in the bacterial DNA (3).
- The non growing population of ofloxacin treated resistant mutants was monitored through flow cytometry.
- The multi drug tolerance of ofloxacin resistant mutants was assessed too.

## APPROACH & PROCEDURE

### Mutant Enrichment Assays

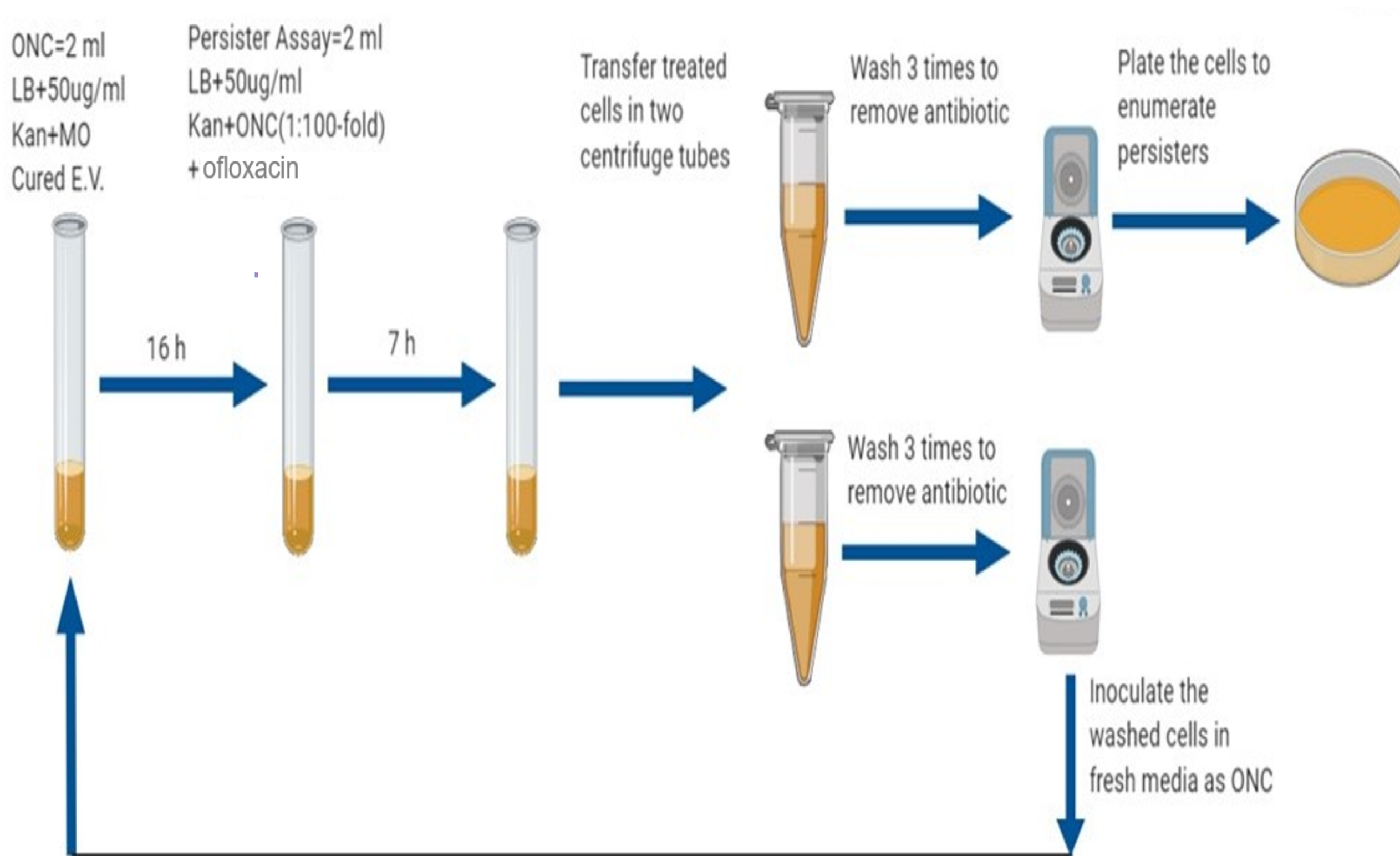


Figure 1. Schematic Diagram of Mutant Enrichment Assay

### Multi Drug Resistance Assays

- Each ofloxacin resistant mutant was treated with gentamicin and ampicillin for 7 hours to assess the multi drug tolerance.

### OD600 Growth Curves

The growth (OD600) of those resistant mutants was observed through a plate reader at designated time points.

### Flow Cytometry

- Flow Cytometric measurements were taken at T=0, T=2, and T=3 of overnight cultures.
- This was used to see the independent growth of each sample by measuring the concentration of mCherry light emission.

## RESULTS & DISCUSSION

- During the 22 day experiment we had 8 out of 10 sample survive the reoccurring antibiotic treatment. (Figure 2)
- All of the samples gained resistance to high concentrations of ofloxacin. (Figure 3)
- 5 samples, (F-J), have multi resistance to both ofloxacin and ampicillin. Sample I is resistant to all three antibiotics that were tested: Ofloxacin, ampicillin, and gentamicin. (Figure 4)

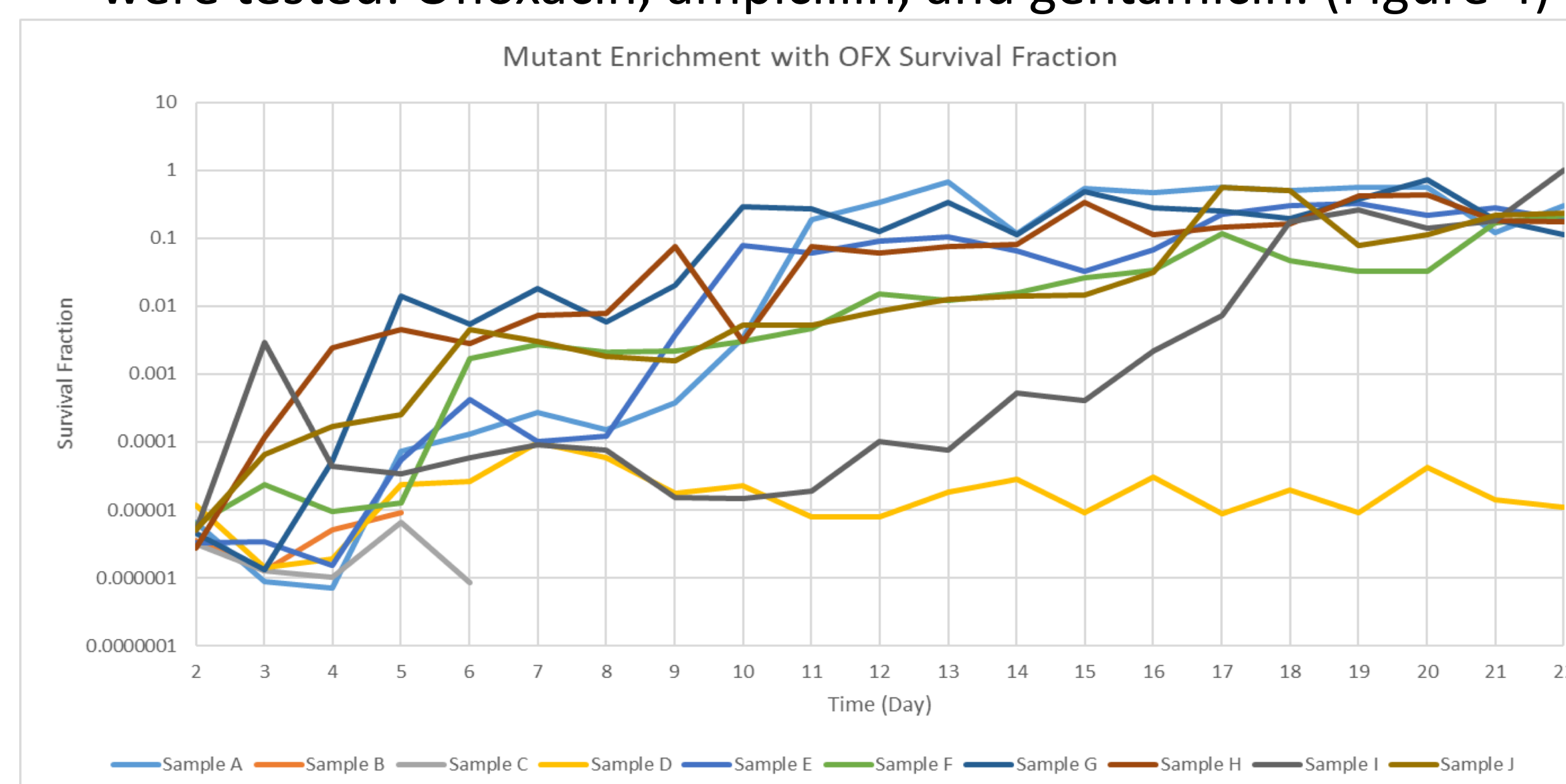


Figure 2. Survival Fraction of resistant mutants emerging from persisters after being treated with Ofloxacin for 22 days.

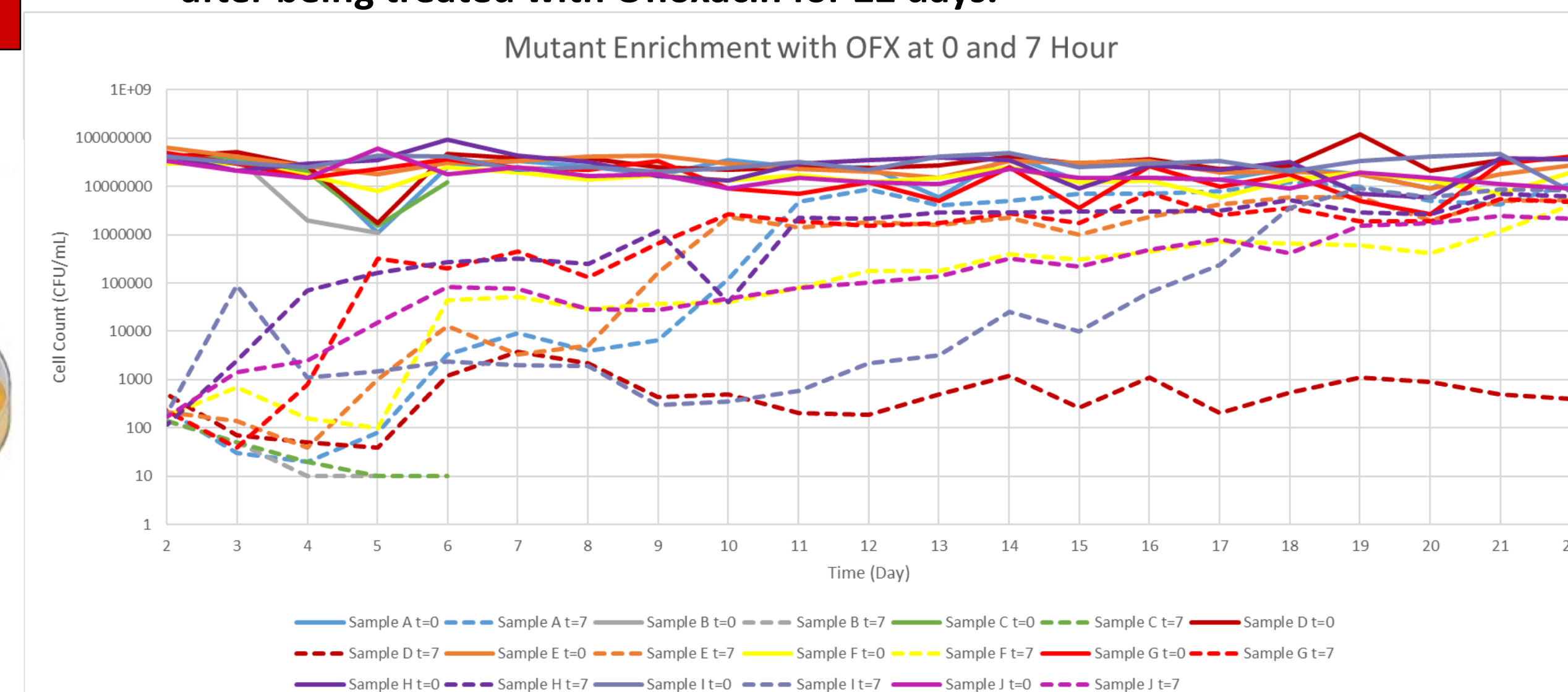


Figure 3. Cell count of resistant mutants emerging from persisters after being treated with Ofloxacin for 22 days at 0<sup>th</sup> hour (Filled line) and 7<sup>th</sup> hour (Dashed Line)

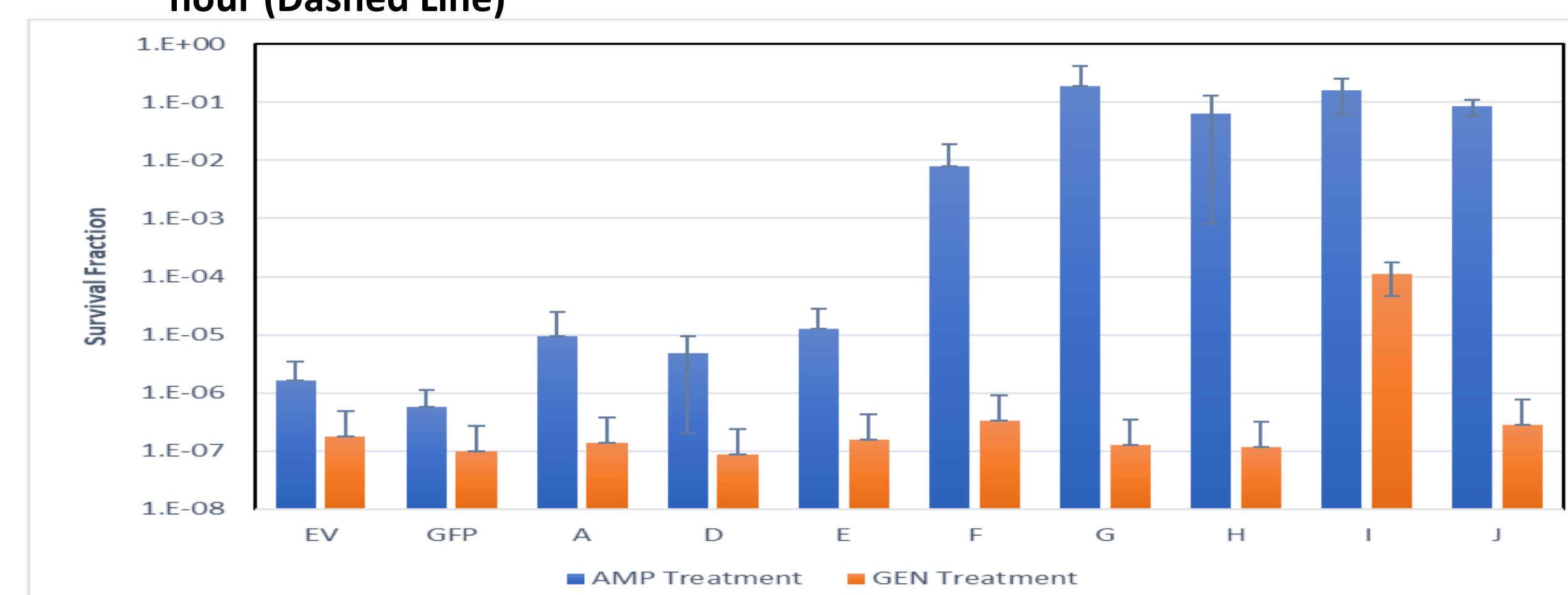


Figure 4. The Data represents the survival fraction of 4 biological replicates for Ampicillin and 3 biological replicates for Gentamicin of 7 hour antibiotic treatment.

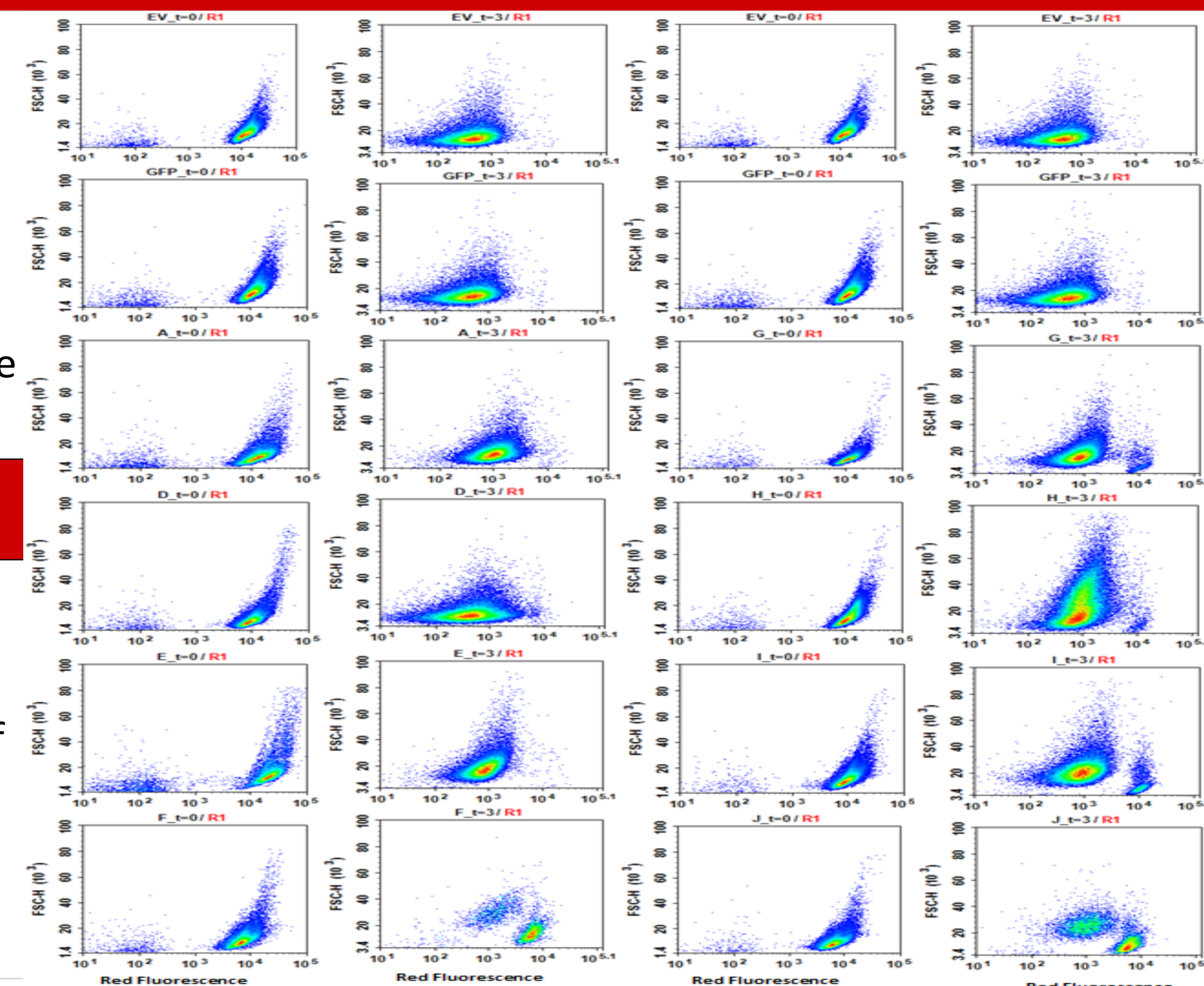


Figure 5. The data represents one of the three biological replicates of the non growing cell assay.

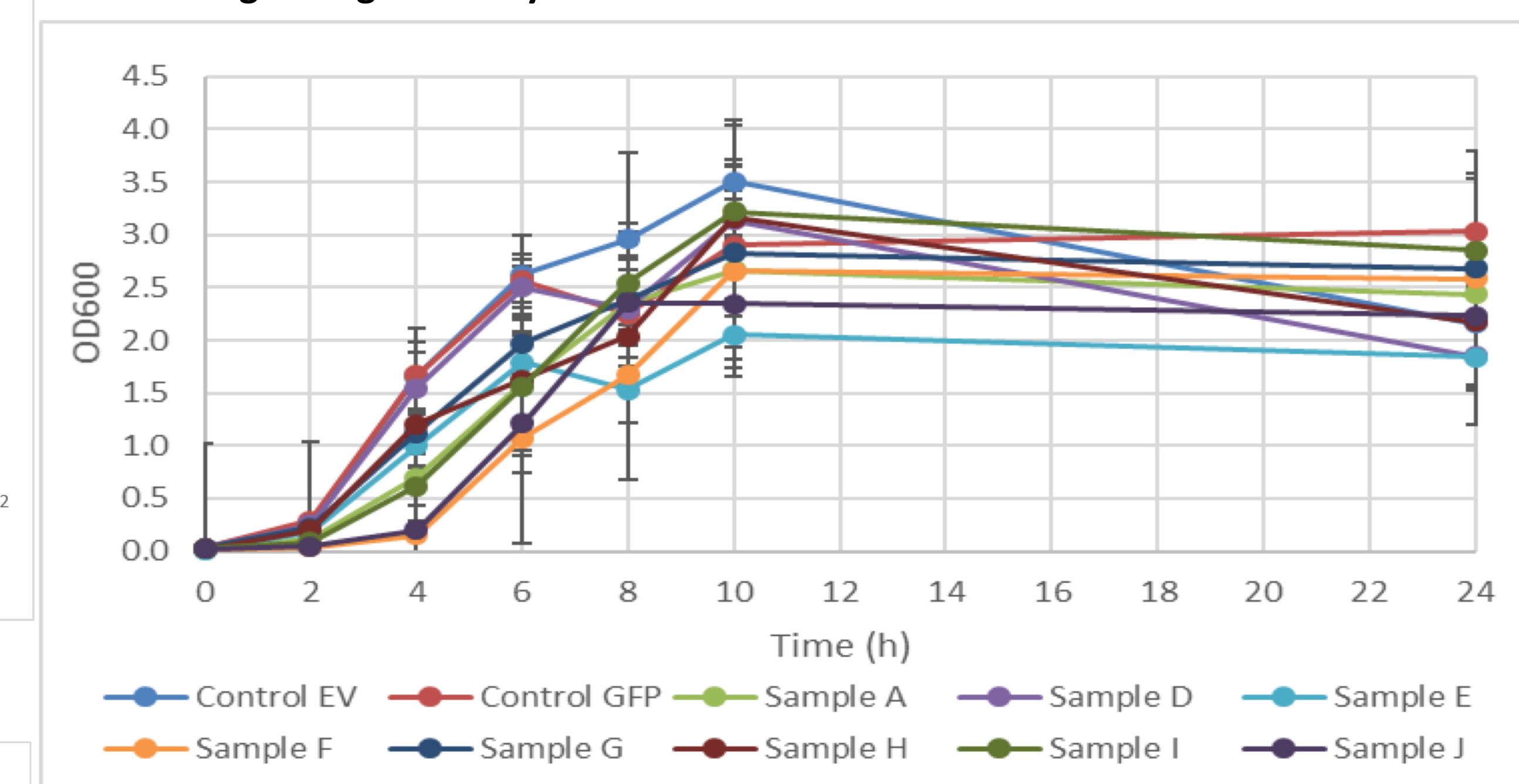


Figure 6. The data represents three biological replicates of 24 hour optic density measurements of two controls and mutant samples.

## FUTURE DIRECTION

Sequencing the genomic DNA of the resistant mutants to identify the possible random mutations that are causing the antibiotic resistance.

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## REFERENCES

- Orman, Mehmet A., and Mark P. Brynildsen. "Dormancy is not necessary or sufficient for bacterial persistence." *Antimicrobial agents and chemotherapy* 57, no. 7 (2013): 3230- 3239.
- Van den Bergh, B., Fauvart, M. and Michiels, J., 2017. Formation, physiology, ecology, evolution and clinical importance of bacterial persisters. *FEMS microbiology reviews*, 41(3), pp.219-251.
- Windels, E. M., Michiels, J. E., Fauvart, M., Wenseleers, T., Bergh, B. V. D., & Michiels, J. (2019). Bacterial persistence promotes the evolution of antibiotic resistance by increasing survival and mutation rates. *The ISME Journal*, 13(5), 1239-1251. doi: 10.1038/s41396-019-0344-9